## **Instructions for Cutting Paraffin Sections**

## **MATERIALS NEEDED:**

- ✓ Treated microscope slides (Silane, Poly-L-lysine) or positive charge ("Plus") slides.
- ✓ **Slide box** to hold slides with sections
- ✓ **Artist paint brushes**. The lab has some available to use, but you may want to purchase your own.
- **✓** Paraffin embedded samples.
- 1. Set water bath to **44°C.** This can be set higher if tissue sections are thicker, or lower if sections break apart or the tissue is fragile.
- 2. Cut the block edges with a razor blade into a trapezoid to help sections cut more easily and help with section orientation.
- 3. Using the **coarse retraction button**, move arm on the microtome <u>ALL</u> the way back, **LOCK HANDLE** and insert block.
- 4. Use the left lever to loosen the blade holder and move blade close to, but not touching, the block. Tighten the left lever.
- 5. The blade is **VERY SHARP**. Avoid touching it with your fingers.
- Set to "trim", cut 30 μm sections until you start cutting into the tissue. Adjust block angle as needed. NOTE: Be sure to LOCK HANDLE to remove block, replace block or when doing any additional trimming with razor blade while on the microtome.
- 7. Remove tissue block and put into the water bath for a few seconds to rehydrate tissue, pat dry, and then put in the -20°C for ~5 min to harden paraffin.
- 8. Label charged ('Plus') or 'silaned' slides with a #2 pencil or histology marker while block is cooling.
- 9. Set microtome to cut ('Feed') **5 μm** sections (or desired thickness).
- 10. Cut a long ribbon (as long as things go well) with enough sections for your intended number of slides.
- 11. Use paint brush to move ribbon to room temperature **50%** ethanol (this helps spreading of tissue).
- 12. Use razor blade to cut ribbon into the desired number of sections for each slide.
- 13. Pick sections up with a large untreated slide, move to warm water bath (set to **44°C**), and let them float.
- 14. Pick up section by moving a labeled, treated slide underneath and lifting it up into the tissue <u>at an angle</u> (this requires practice). **NOTE: you <u>cannot</u> reposition sections once they adhere to the slides!**
- 15. Tap off excess water and dry bottom of slide, place slide in rack to drain and then move slides to slide warmer (set at ~44°C).
- 16. Leave overnight on slide warmer to dry.